

A SYSTEMATIC ANALYSIS OF ANTIBIOTICS USING PAPER CHROMATOGRAPHY

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INTRODUCTION

Paper chromatographic studies of compounds in several solvent systems for the purpose of their classification and identification have proved useful in systematic analysis¹. At a recent symposium² papers were presented on the systematic analysis of alkaloids³, antibiotics⁴, synthetic dyes⁵, inorganic cations and anions^{6,7}, aromatic compounds⁸ and steroids⁹.

Several methods for the systematic analysis of antibiotics are known. The oldest of them is the so-called "summarized paper chromatogram"¹⁰, where the antibiotics are analysed in eight solvent systems and the R_F values of an antibiotic are presented graphically as a "summarized chromatogram". This principle has been used with various modifications by many authors¹¹⁻¹⁷.

MIYAZAKI *et al.*¹⁸ classified antibiotics using a salting-out paper chromatography technique, in which nine solvent systems with different concentrations of ammonium chloride in water were used. According to their R_F values in these systems antibiotics can be divided into six groups^{18,19}. NEMEC *et al.*²⁰ have also applied it to antibiotics from fungi.

BETINA²¹ and BETINA AND NEMEC²² determined the ionic character of unknown antibiotics and also the possibility of their isolation and purification from the "pH chromatograms".

From the chemical point of view antibiotics are a very heterogeneous group of biologically active compounds, which causes some difficulty in working out a systematic chromatographic analysis. Bearing this fact in mind a systematic analysis was developed along the following lines. The antibiotics are first analysed in four solvent systems, and then are divided into five classes with fourteen subclasses which are further analysed in additional solvent systems for each class.

MATERIALS AND METHODS

Principal solvent systems

The antibiotics were first analysed in the following four solvent systems:

1. Distilled water,
2. *n*-Butanol saturated with water,
3. Ethyl acetate saturated with water,
4. Benzene saturated with water.

The antibiotics were divided into five classes with fourteen subclasses, according to their R_F values in the above solvent systems.

*Additional solvent systems**

The following additional solvent systems were used for individual classes of antibiotics:

Class I:

- A. Methanol-water (40:60).
- B. *n*-Propanol-water (40:60).
- C. Methanol-3 % ammonium chloride in water (70:30).
- D. Methyl ethyl ketone-*n*-butanol-water (30:5:65).

Classes II and III**:

- E. 3 % Ammonium chloride in water.
- F. Isoamyl acetate-methanol-99 % formic acid-water (65:20:5:10).
- G. *n*-Butyl acetate-methyl ethyl ketone-0.15 *M* phosphate buffer, pH 7.4 (50:25:5).
- H. Ethyl acetate-*n*-hexane-0.15 *M* phosphate buffer, pH 6.0 (65:15:20).

Classes IV and V***:

- I. Isoamyl acetate-methanol-99 % formic acid-water (40:20:10:30).
- J. *n*-Butanol-methanol-water (40:10:50).
- K. Methanol-*n*-hexane (60:40).
- L. Benzene-cyclohexanone-0.15 *M* phosphate buffer, pH 7.4 (5:35:60).

Development and detection of chromatograms

Strips of the Whatman No. 1 paper 1×35 cm were used, the origin being 3 cm from the lower end of the strips.

For the development of the chromatograms 500 ml glass cylinders were used with 25 ml of the solvents. After applying the antibiotics to the origin and drying in air, the chromatographic strips were immersed to a depth of 1 cm in the solvent. Ascending development at $20 \pm 1^\circ$, without preliminary saturation of the chromatograms with the vapours of the solvents, was used. The development was stopped when the solvent front reached a distance of 15 cm from the origin.

After drying the chromatograms in air, detection was carried out either bioautographically, using sensitive test microorganisms²³, or, in some special cases, chemically (see Table I).

Antibiotics

Sixty-two antibiotics from actinomycetes, fungi and lichens were studied. Data about the amount of each antibiotic used, the solvent for its solution before application on to paper strips, and the detection technique are given in Table I.

* All proportions are given by volume.

** Upper layers of systems F, G, and H were used.

*** Bottom layers of systems I, J, K, and L were used.

TABLE I

SOLVENTS AND DETECTION METHODS FOR THE ANTIBIOTICS ANALYZED

Solvents used for the application of antibiotics on to chromatograms: A = acetone; AW = 50% acetone in water; C = chloroform; E = ethyl acetate; M = methanol; W = water. Detection methods: a = bioautographically with *Bacillus subtilis*; b = bioautographically with *Escherichia coli*; c = bioautographically with *Candida pseudotropicalis*; d = 3% FeCl₃ in methanol; e = 3% NH₄Cl in water; f = acidic KMnO₄ (according to ref. 25, p. 737); g = conc. H₂SO₄; h = 3% NH₄OH in water; i = original colour.

| Antibiotic | Application | | Detection | Colour reaction |
|----------------------------------|-------------------|---------|-----------|-----------------|
| | Amount (μ g) | Solvent | | |
| Aburamycin | 20 | M | a | — |
| Actinomycin C·HCl | 20 | A | a | — |
| Alternariol | 60 | A | d | dark green |
| Amphomycin | 40 | W | a | — |
| Amphotericin B | 250 | W | g | blue |
| Aspergillilic acid | 60 | A | d | red |
| Azalomycin B | 80 | M | a | — |
| Azalomycin F | 60 | M | c | — |
| Bryamycin | 40 | C | a | — |
| Chloramphenicol | 10 | M | a | — |
| Chlorotetracycline·HCl | 30 | M | d | light brown |
| Citrinin | 80 | A | a | — |
| Congocidin·HCl | 40 | W | a | — |
| Cyanein | 80 | A | c | — |
| Cyclopaldic acid | 40 | A | d | dark green |
| Cycloserine | 40 | W | a | — |
| Dihydrostreptomycin (sulfate) | 10 | W | b | — |
| Eniantin B | 90 | A | g | light red |
| Erythromycin (lactobionate) | 20 | W | a | — |
| Etamycin | 5 | A | a | — |
| Flavipin | 50 | M | d | dark green |
| Flavofungin | 80 | M | f | decolorized |
| Fomecin A | 30 | A | d | blue-green |
| Frequentin | 50 | C | d | brown-red |
| Fuscin | 40 | A | h | violet |
| Geodin | 30 | A | a | — |
| Gladiolic acid | 50 | A | e | dark green |
| Gliotoxin | 30 | A | a | — |
| Griseofulvin | 100 | C | g | yellow |
| Illudin M | 40 | A | a | — |
| Illudin S | 20 | A | a | — |
| Javanicin | 20 | A | i | red-violet |
| Kanamycin (sulfate) | 80 | W | b | — |
| Kojic acid | 40 | A | d | brown-red |
| 5-Methoxy- <i>p</i> -toluquinone | 40 | A | a | — |
| Mycophenolic acid | 80 | A | d | dark green |
| Neomycin B (base) | 250 | W | b | — |
| Novobiocin (Na salt) | 10 | E | a | — |

(Table continued on p. 382)

TABLE I (continued)

| Antibiotic | Application | | Detection | Colour reaction |
|--------------------------------|-------------------|---------|-----------|-----------------|
| | Amount (μ g) | Solvent | | |
| Oleandomycin (base) | 30 | M | a | — |
| Oxytetracycline·HCl | 60 | W | d | light brown |
| Patulin | 100 | A | a | — |
| Penicillic acid | 160 | M | a | — |
| Penicillin G (K salt) | 0.5* | W | a | — |
| Penicillin V (K salt) | 0.5* | W | a | — |
| Pleuromutilin | 120 | A | a | — |
| Puromycin | 120 | W | a | — |
| Quadrilineatin | 40 | A | e | dark green |
| Ristocetin | 40 | W | a | — |
| Rugulosin | 20 | A | d | yellow-brown |
| Sclerotiorin | 20 | A | g | orange-red |
| Spinulosin | 50 | A | d | brown |
| Spiramycin | 80 | M | a | — |
| Streptomycin (sulfate) | 10 | W | b | — |
| Synnematin B (cephalosporin N) | 10 | W | a | — |
| Telomycin | 40 | AW | a | — |
| Terreic acid | 80 | M | d | brown-red |
| Tetracycline·HCl | 60 | W | d | light brown |
| Trichothecin | 20 | A | c | — |
| Usnic acid | 80 | A | g | brown |
| Vancomycin | 20 | W | a | — |
| Viomycin (sulphate) | 160 | W | b | — |
| Viridin | 80 | A | a | — |

* Units

RESULTS

The principles whereby the antibiotics were divided into five classes with fourteen subclasses, according to their R_F values in four solvent systems, are given schematically in Table II. Each subclass is represented by a typical antibiotic in Fig. 1.

The distribution of the antibiotics studied into the classes and subclasses is given in Tables III to VII. R_F values in the four principal and in four of the additional solvent systems are presented graphically as "summarized chromatograms" (or " R_F spectra") in Figs. 2 to 6.

Class I of antibiotics is characterized as follows: R_F values are equal to zero in the principal solvent systems 3 and 4, and 0.00–0.30 in system 2. There are greater differences in system 1, these were used for the division of this class into three subclasses: 1a, 1b, and 1c (see Table II and Fig. 1).

Class I includes antibiotics of the streptomycin–neomycin group, tetracyclines, cycloserine, synnematin B (cephalosporin N), vancomycin, ristocetin and other antibiotics (Table III). The "summarized chromatograms" of tetracyclines are very similar. There are also similarities between the "summarized chromatograms" of

TABLE II

CLASSIFICATION OF ANTIBIOTICS INTO CLASSES (PART A) AND INTO SUBCLASSES (PARTS B, C, D, E AND F)

| <i>Part A</i> | | <i>Classes</i> | | | | |
|--|-----------|----------------|-------------------------|-------------------------|-----------|--|
| <i>R_F</i> values in principal systems | <i>I</i> | <i>II</i> | <i>III</i> | <i>IV</i> | <i>V</i> | |
| <i>R_{F1}</i> | > 0.00 | > 0.60 | 0.31-0.60 | 0.00-0.30 | 0.00-1.00 | |
| <i>R_{F2}</i> | 0.00-0.30 | > 0.30 | > <i>R_{F1}</i> | > <i>R_{F1}</i> | > 0.60 | |
| <i>R_{F3}</i> | 0.00 | > 0.00 | > <i>R_{F4}</i> | ≅ <i>R_{F4}</i> | > 0.60 | |
| <i>R_{F4}</i> | 0.00 | 0.00 | 0.00-0.60 | 0.00-0.60 | > 0.60 | |

| <i>Part B</i> | | <i>Class I</i> | | |
|-----------------------------|-------------------|----------------|-----------|--|
| <i>R_F</i> values | <i>Subclasses</i> | | | |
| | <i>Ia</i> | <i>Ib</i> | <i>Ic</i> | |
| <i>R_{F1}</i> | > 0.60 | 0.31-0.60 | ≤ 0.30 | |

| <i>Part C</i> | | <i>Class II</i> | | |
|-----------------------------|-------------------------|-------------------------|-------------------------|--|
| <i>R_F</i> values | <i>Subclasses</i> | | | |
| | <i>IIa</i> | <i>IIb</i> | <i>IIc</i> | |
| <i>R_{F1}</i> | > <i>R_{F2}</i> | > <i>R_{F2}</i> | < <i>R_{F3}</i> | |
| <i>R_{F2}</i> | > <i>R_{F1}</i> | < <i>R_{F2}</i> | < <i>R_{F3}</i> | |
| <i>R_{F3}</i> | > <i>R_{F1}</i> | < <i>R_{F2}</i> | < <i>R_{F3}</i> | |

| <i>Part D</i> | | <i>Class III</i> | |
|-----------------------------|-------------------|------------------|--|
| <i>R_F</i> values | <i>Subclasses</i> | | |
| | <i>IIIa</i> | <i>IIIb</i> | |
| <i>R_{F4}</i> | 0.00 | 0.05-0.60 | |

| <i>Part E</i> | | <i>Class IV</i> | | |
|-----------------------------|-------------------------|-------------------------|-------------------------|--|
| <i>R_F</i> values | <i>Subclasses</i> | | | |
| | <i>IVa</i> | <i>IVb</i> | <i>IVc</i> | |
| <i>R_{F2}</i> | > <i>R_{F1}</i> | > <i>R_{F1}</i> | > <i>R_{F1}</i> | |
| <i>R_{F3}</i> | > <i>R_{F1}</i> | 0.00 | > <i>R_{F4}</i> | |
| <i>R_{F4}</i> | 0.00 | 0.00 | 0.05-0.60 | |

| <i>Part F</i> | | <i>Class V</i> | | |
|-----------------------------|-------------------|----------------|-----------|--|
| <i>R_F</i> values | <i>Subclasses</i> | | | |
| | <i>Va</i> | <i>Vb</i> | <i>Vc</i> | |
| <i>R_{F1}</i> | > 0.60 | 0.31-0.60 | 0.00-0.30 | |

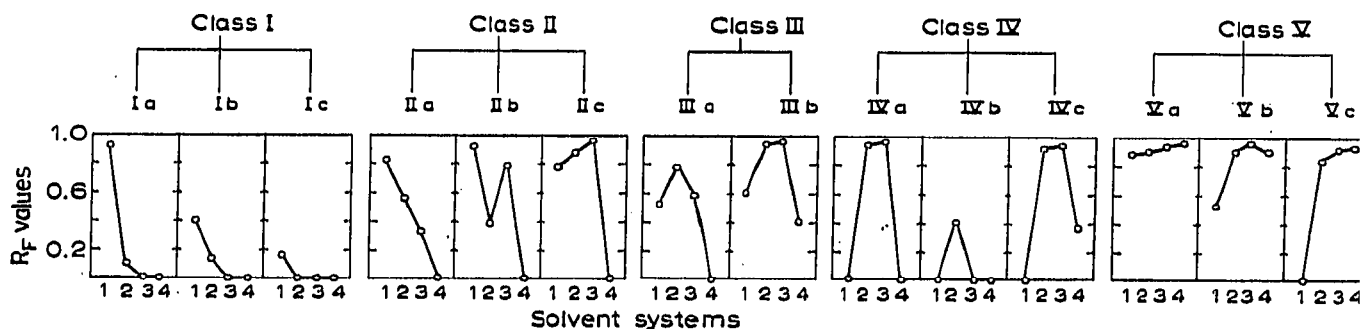


Fig. 1. Typical antibiotics belonging to five classes according to their R_F values in principal solvent systems 1, 2, 3 and 4. From left to right: cycloserine (subclass Ia), chlorotetracycline (Ib) streptomycin (Ic), kojic acid (IIa), penicillin G (IIb), chloramphenicol (IIc), oleandomycin (IIIa) etamycin (IIIb), cyanein (IVa), azalomycin F (IVb), azalomycin B (IVc), illudin M (Va), pleuro mutilin (Vb) and viridin (Vc).

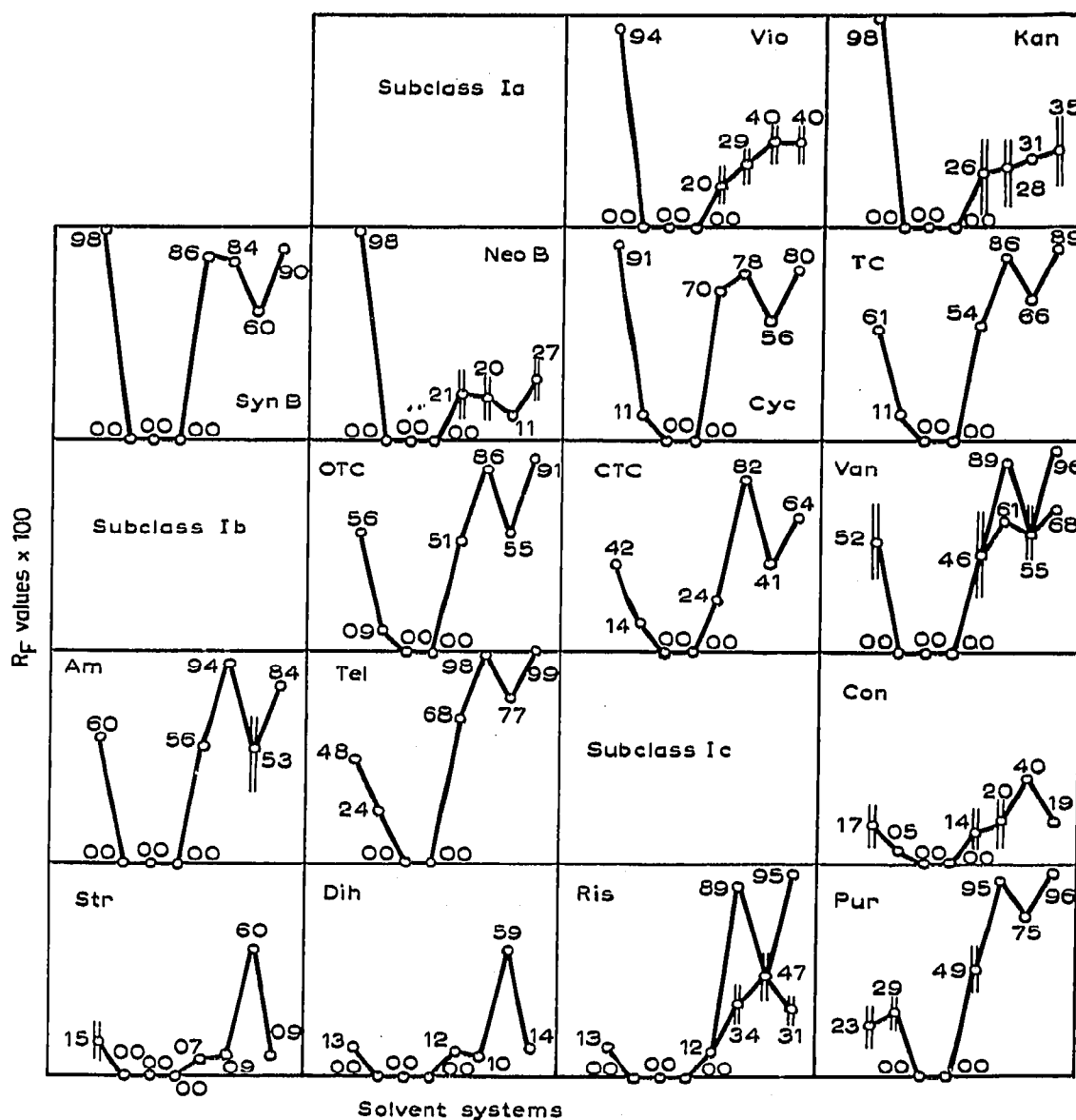


Fig. 2. "Summarized chromatograms" of antibiotics of class I. Subclass Ia: viomycin (Vio), kanamycin (Kan), synnematin B (Syn B), neomycin B (Neo B), cycloserine (Cyc) and tetracycline (TC). Subclass Ib: oxytetracycline (OTC), chlorotetracycline (CTC), vancomycin (Van), amphomycin (Am) and telomycin (Tel). Subclass Ic: congocidin (Con), streptomycin (Str), dihydrostreptomycin (Dih), ristocetin (Ris) and puromycin (Pur). Solvent systems: 1, 2, 3, 4, A, B, C, and D.

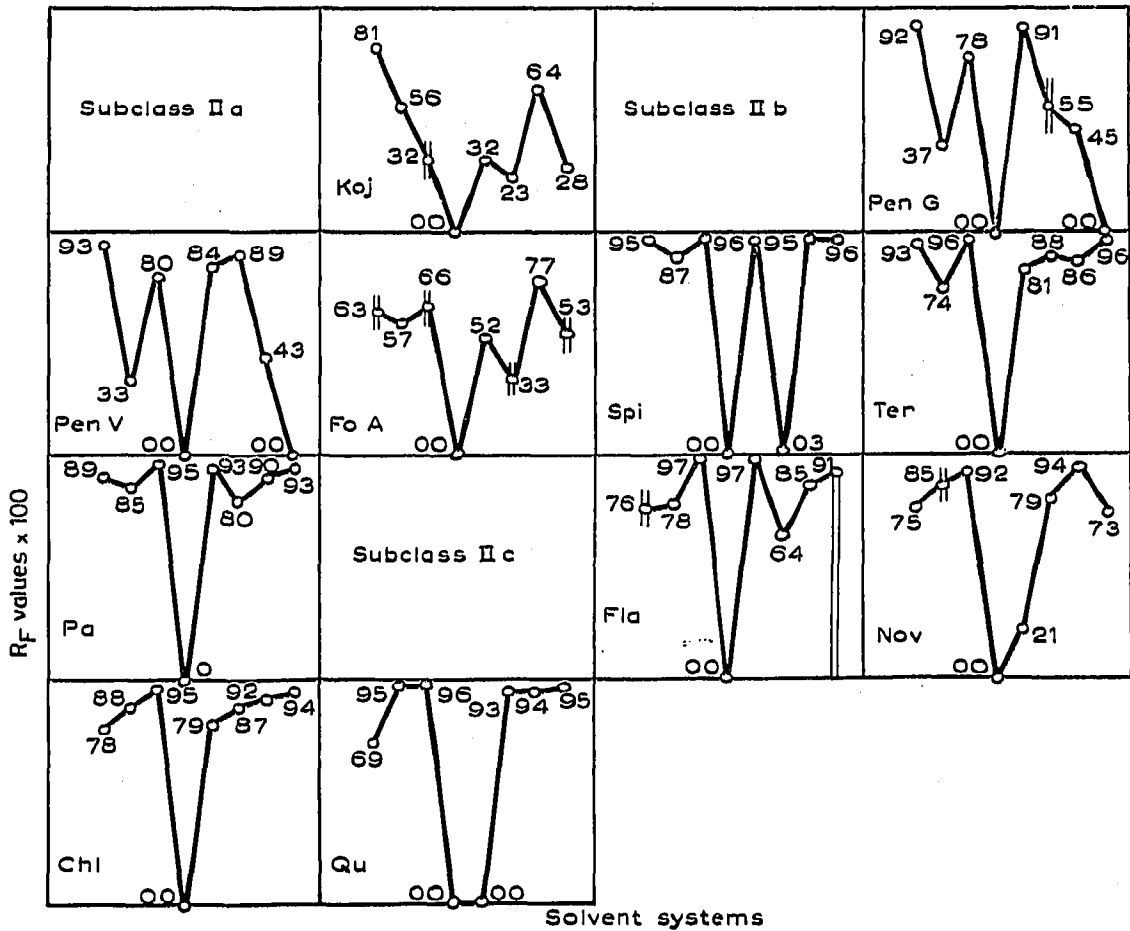


Fig. 3. "Summarized chromatograms" of antibiotics of class II. Subclass IIa: kojic acid (Koj). Subclass IIb: penicillin G (Pen G), penicillin V (Pen V), fomecin A (Fo A), spiramycin (Spi), terreic acid (Ter) and penicillic acid (Pa). Subclass IIc: flavipin (Fla), novobiocin (Nov), chloramphenicol (Chl) and quadrilincatin (Qu). Solvent systems: 1, 2, 3, 4, E, F, G, and H.

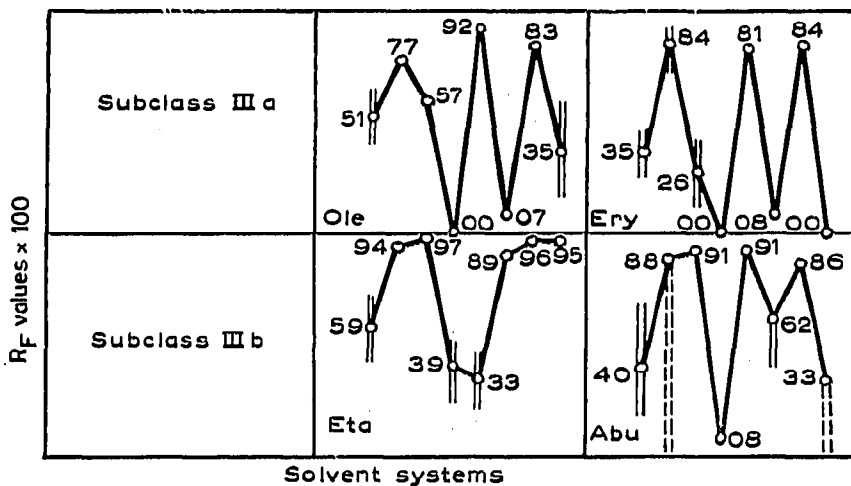


Fig. 4. "Summarized chromatograms" of antibiotics of class III. Subclass IIIa: oleandomycin (Ole) and erythromycin (Ery). Subclass IIIb: etamycin (Eta) and aburamycin (Abu). Solvent systems: 1, 2, 3, 4, E, F, G, and H.

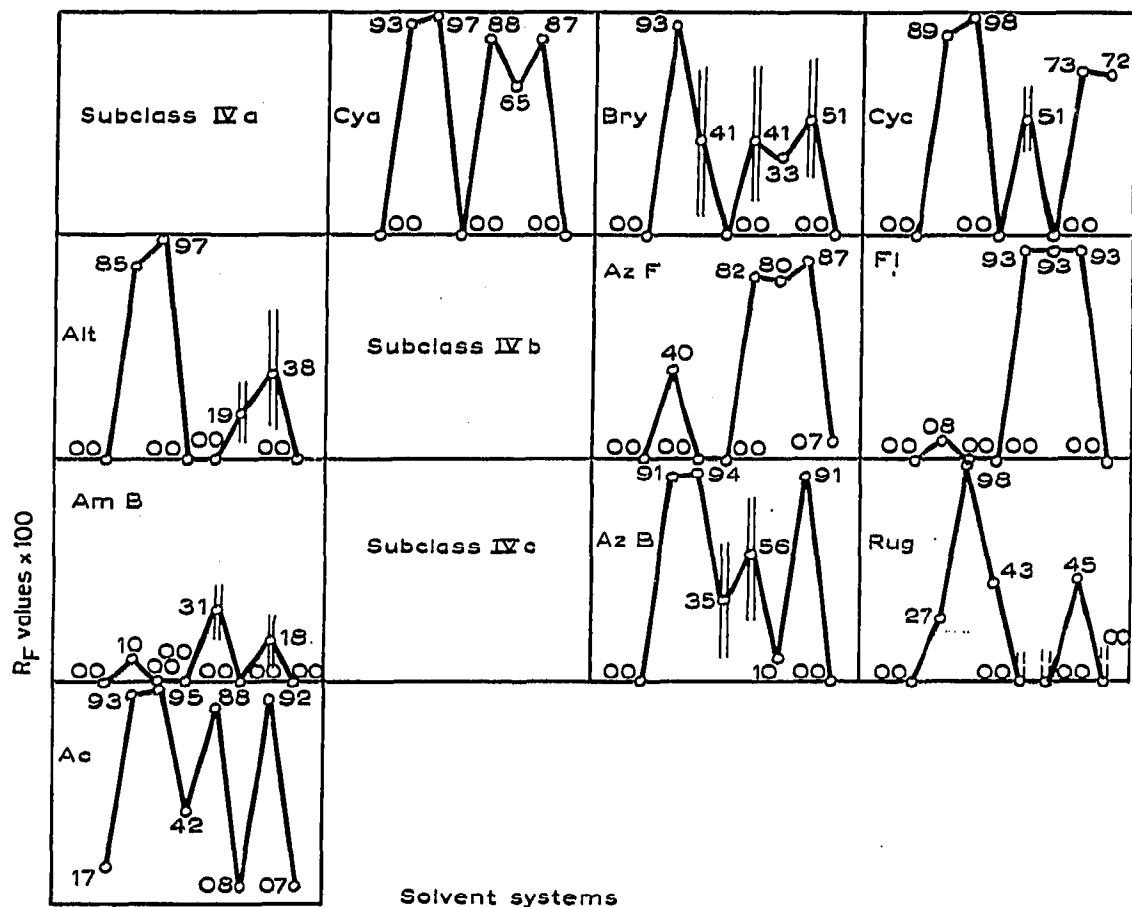


Fig. 5. "Summarized chromatograms" of antibiotics of class IV. Subclass IVa: cyancin (Cya), bryamycin (Bry), cyclopaldic acid (Cyc) and alternariol (Alt). Subclass IVb: azalomycin F (Az F), flavofungin (Fl) and amphotericin B (Am B). Subclass IVc: azalomycin B (Az B), rugulosin (Rug) and actinomycin C (Ac). Solvent systems: 1, 2, 3, 4, I, J, K, and L.

TABLE III
ANTIBIOTICS BELONGING TO CLASS I

| Subclass Ia | Subclass Ib | Subclass Ic |
|--------------|--------------------|---------------------|
| Viomycin | Oxytetracycline | Streptomycin |
| Kanamycin | Chlorotetracycline | Dihydrostreptomycin |
| Synnematin B | Vancomycin | Congocidin |
| Neomycin B | Amphomycin | Ristocetin |
| Cycloserine | Telomycin | Puromycin |
| Tetracycline | | |

TABLE IV
ANTIBIOTICS BELONGING TO CLASS II

| Subclass IIa | Subclass IIb | Subclass IIc |
|--------------|-----------------|-----------------|
| Kojic acid | Penicillin G | Flavipin |
| | Penicillin V | Novobiocin |
| | Fomecin A | Chloramphenicol |
| | Spiramycin | Quadrilineatin |
| | Terreic acid | |
| | Penicillic acid | |

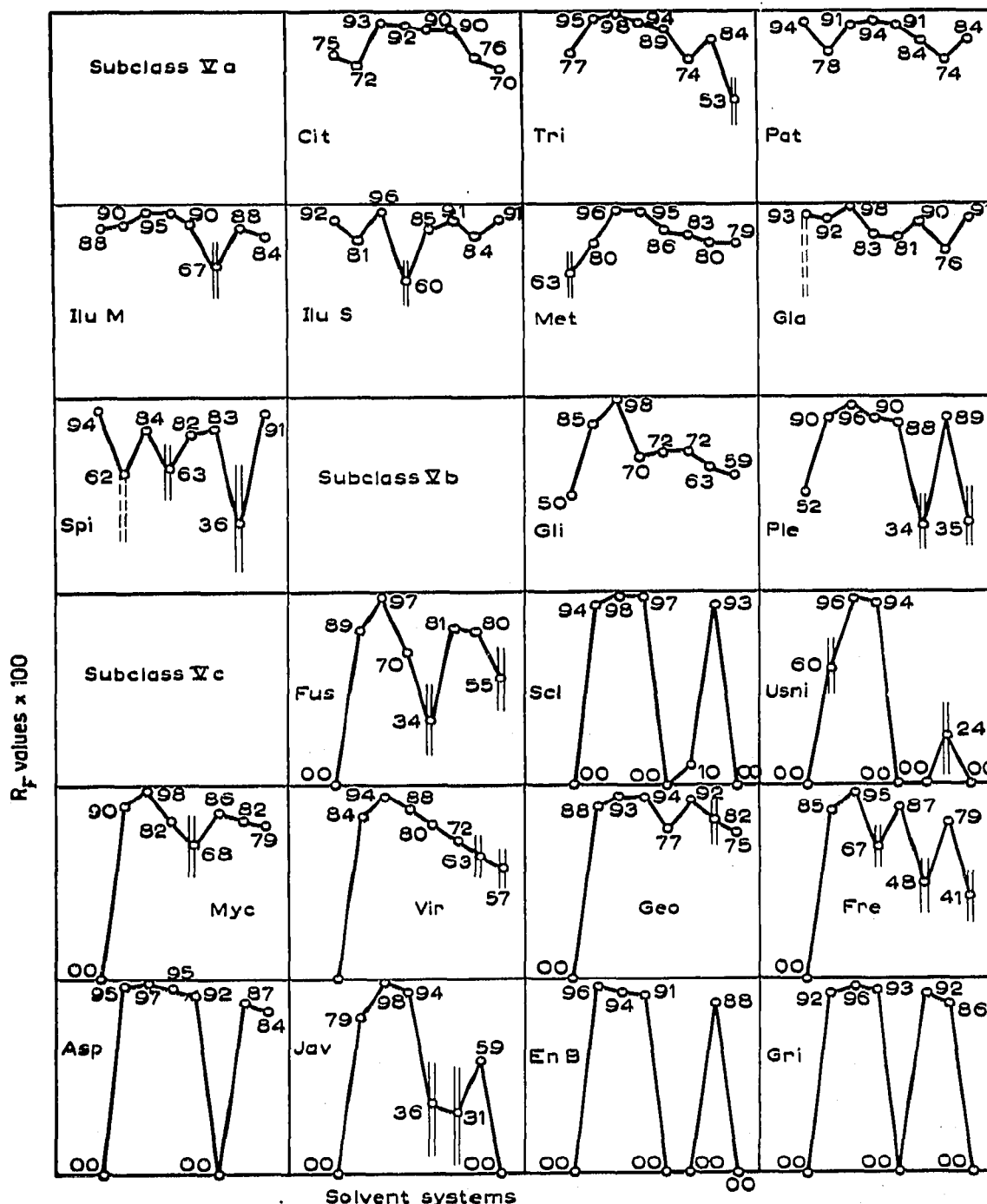


Fig. 6. "Summarized chromatograms" of antibiotics of class V. Subclass Va: citrinin (Cit), trichothecin (Tri), patulin (Pat), illudin M (Ilu M), illudin S (Ilu S), 5-methoxy-*p*-toluquinone (Met), gladiolic acid (Gla) and spinulosin (Spi). Subclass Vb: gliotoxin (Gli) and pleuromutilin (Ple). Subclass Vc: fusicin (Fus), sclerotiorin (Scl), usnic acid (Usni), mycophenolic acid (Myc), viridin (Vir), geodin (Geo), frequentin (Fre), aspergillilic acid (Asp), javanicin (Jav), eniantin B (En B) and griseofulvin (Gri). Solvent systems: 1, 2, 3, 4, I, J, K, and L.

streptomycin and dihydrostreptomycin. Commercial samples of vancomycin (Vanco-cin) and ristocetin (Spontin) were used in our studies. These preparations contain mixtures of vancomycins and of ristocetins, respectively. Double spots in systems

TABLE V
ANTIBIOTICS BELONGING TO CLASS III

| <i>Subclass III a</i> | <i>Subclass III b</i> |
|------------------------------|------------------------|
| Oleandomycin Erythromycin | Etamycin Aburamycin |

TABLE VI
ANTIBIOTICS BELONGING TO CLASS IV

| <i>Subclass IV a</i> | <i>Subclass IV b</i> | <i>Subclass IV c</i> |
|---|---|--|
| Cyanein Bryamycin Cyclopaldic acid Alternariol | Azalomycin F Flavofungin Amphotericin B | Azalomycin B Rugulosin Actinomycin C |

TABLE VII
ANTIBIOTICS BELONGING TO CLASS V

| <i>Subclass V a</i> | <i>Subclass V b</i> | <i>Subclass V c</i> |
|--|----------------------------|---|
| Citrinin Trichothecin Patulin Illudin M Illudin S 5-Methoxy- <i>p</i> -toluquinone Gladiolic acid Spinulosin | Gliotoxin Pleuromutilin | Fuscin Sclerotiorin Usnic acid Mycophenolic acid Viridin Geodin Frequentin Aspergilliacid Javanicin Eniantin B Griseofulvin |

B and D were observed in both cases; these solvent systems could be used for the paper chromatographic separation of the antibiotics mentioned.

Class II of antibiotics differ from class I with respect to their R_F values in the solvent systems 2, 3 and partially in system 1 (see Table II and Fig. 1). Differences in the relationships of the R_F values in systems 1, 2, and 3 served as limits for the subclasses II a, II b and II c.

Class III of antibiotics differ from class II by the R_F values in solvent system 1 and partially (subclass III b) in solvent system 4. Differences of R_F values in system 4 were used for the characterization of subclasses III a and III b.

Both class II and class III have the same additional solvent systems because of some similarities in antibiotics belonging to these two classes. Such antibiotics as penicillins G and V, chloramphenicol, novobiocin and macrolide antibiotics can be shown in these classes. Against this, synnematin B (cephalosporin N), a penicillin with a more hydrophilic side chain in its molecule, does not belong to class II like

penicillins G and V, but to class I which includes antibiotics of a more hydrophilic character.

Antibiotics of class IV have some similarities with classes I, II, and III, when their R_F values in solvent systems 2, 3 and 4 are considered. However, they are akin to the Vc subclass by virtue of their R_F values in solvent system 1. They form a transitional class in our systematic analysis.

Class V is characterized by the R_F values in systems 2, 3 and 4, which are always higher than 0.60, showing a maximum in system 3 in most cases. The characterizations of subclasses Va, Vb, and Vc were established according to differences of the R_F values in solvent system 1.

DISCUSSION

DROZEN²⁴ discussed the classification and identification procedures of systematic chromatographic analysis in the light of information theory. He mentioned two main different methods of analysis, *viz.* the so-called sequential (step-wise) method and the simultaneous method.

In the chromatographic analysis of antibiotics hitherto known, both principles have been used. SNELL *et al.*¹⁴ classified and identified several peptide antibiotics according to the principles of sequential analysis. ISHIDA *et al.*¹⁰ and most other authors used the principles of simultaneous analysis in several solvent systems. Both methods have their advantages and disadvantages²⁴.

The advantage of our method of systematic analysis of antibiotics is the use of a combination of both principles. In the first step, antibiotics are analysed simultaneously in the principal solvent systems and are classified into classes and subclasses. The classification is then completed by the second step using the additional solvent systems. Using our combination of sequential and simultaneous analysis, it is not necessary to analyse a compound in many solvent systems as is done in simultaneous analysis. On the other hand, it is possible to compare the "summarized chromatogram" of an antibiotic with others belonging to the same class or subclass, respectively. The chromatographic identification of an unknown antibiotic, of course, must be confirmed by other data (I. R. spectrum, U. V. spectrum and other physical, chemical and biological properties).

MACEK AND PROCHÁZKA²⁵ mention two possibilities for the evaluation of solvent systems for paper chromatography. Solvent systems are classified according to their decreasing polarity in the eluotropic scale. Another classification of solvent systems is based on their possibility (or impossibility) to be hydrogen donors or acceptors, forming hydrogen bridges.

Both criteria were taken into consideration in our attempt to establish a systematic analysis of antibiotics. The polarity of the principal solvent systems decreases from system 1 to system 4. Solvent system 1 consists of water, which can accept or donate hydrogen, forming hydrogen bridges with other molecules of water. *n*-Butanol, the main component of system 2, also possesses the property of accepting or donating hydrogen and forms hydrogen bridges with molecules of other compounds. Ethyl acetate in solvent system 3 can only accept hydrogen, and benzene in system 4 has the least possibility (compared to the three solvents mentioned) of forming hydrogen bridges.

According to our classification, the hydrophilic character of the antibiotics analysed generally diminishes from subclass Ia to subclass Vc. Structurally related antibiotics (such as tetracyclines, streptomycins, penicillins G and V) have very similar "summarized chromatograms". In some cases the additional solvent systems may help to separate closely related antibiotics (see vancomycin and ristocetin in systems B and D, and penicillins G and V in system F).

In searches for new antibiotics from fungi our systematic chromatographic analysis can be applied when only crude concentrates of the antibiotics are available. An antibiotically active strain is cultivated in liquid medium up to the maximum of the antibiotic activity. Then the mycelium of the fungus is separated from the medium by filtration. Both the filtrate of the cultivation medium and the mycelium of the producing strain are used separately for the preparation of crude concentrates of antibiotics. The crude concentrates are prepared as follows²⁶:

(1) 50 ml of acetone are added to 50 ml of the filtrate to precipitate proteins and other compounds that might interfere in the chromatographic analysis of the antibiotics in the filtrate. The mixture is heated to 50° for 10 min and then cooled to laboratory temperature. After precipitation and filtration, the filtrate is evaporated *in vacuo* to dryness. The dry residue is dissolved in 5 ml of 80 % aqueous acetone, filtered and used for the chromatographic studies.

(2) The mycelium of the producing strain is extracted twice with ethyl acetate, filtered and the filtrate is evaporated *in vacuo* to dryness. The dry residue is dissolved in 5 ml of 80 % aqueous acetone, filtered and used for the chromatographic studies of antibiotics from the mycelium.

Results of such chromatographic studies²⁶ of antibiotics from filtrates of media and from mycelia of 62 strains of antibiotically active fungi are given in Table VIII.

DOSKOČILOVÁ AND VONDRÁČEK¹⁷ in their studies of antibiotics from actinomycetes prepared only concentrates from media and used them for chromatographic analysis of antibiotics. Other authors²⁷ prepared mixed concentrates of antibiotics

TABLE VIII
CLASSIFICATION OF ANTIBIOTICS FROM 62 STRAINS OF FUNGI

| Subclasses | Antibiotics found in | | |
|------------|----------------------|---------|------------------------|
| | Media | Mycelia | Both media and mycelia |
| Ia | 10 | — | — |
| Ib | 1 | — | — |
| Ic | — | — | — |
| IIa | 5 | — | — |
| IIb | 1 | — | — |
| IIc | 5 | — | — |
| IIIa | — | — | — |
| IIIb | — | 1 | — |
| IVa | 5 | 5 | 5 |
| IVb | 7 | — | — |
| IVc | — | 1 | — |
| Va | 1 | 6 | 3 |
| Vb | — | — | — |
| Vc | — | 28 | 1 |
| | 35 | 41 | 9 |

from both the cultivation media and the mycelia of fungi and used them for chromatographic studies.

We found that it is necessary to prepare concentrates from both the cultivation media and the mycelia of strains studied, and that a separate chromatographic analysis of both concentrates of the same strain provides more precise results. We found in many cases that antibiotics in the concentrate from the medium and in the concentrate from the mycelium of the same strain belonged to different classes or subclasses of our systematic analysis. In some cases it was also possible to identify chromatographically antibiotics in concentrates from the medium and from the mycelium of the same strain. Such findings are also very useful for isolation procedures of unknown antibiotics.

In our laboratory data of these chromatographic studies are used not only for the classification but also for the tentative identification of unknown antibiotics. In our screening programme of new antibiotics from fungi it was possible to identify three antibiotics which had been isolated as strictly identical with citrinin²⁸, gliotoxin²⁹ and kojic acid²⁹. Chromatographic identifications (the "summarized chromatograms", the "pH chromatograms" and the salting-out paper chromatograms) were confirmed by the I. R. spectra and by other physical, chemical and biological properties of these antibiotics.

Data about antibiotics in crude concentrates from filtrates of cultivation media and from mycelia of fungi that are obtained from the "summarized chromatograms" and from the "pH chromatograms"^{21, 22} help us to determine their ionic character and general possibilities of the isolation procedures^{21, 22, 29}. This principle was applied during isolation and purification procedures of the three antibiotics mentioned and of the new antibiotic cyanein³⁰.

Of the known antibiotics studied, antibacterial compounds belong mainly to classes I, II and III, whereas antifungal antibiotics or antibiotics with simultaneous antibacterial and antifungal activity belong mostly to classes IV and V, respectively. It is well known that solubility, distribution coefficients and other physico-chemical properties of antibiotics have some relationship to their spectra of antimicrobial activity. This aspect of our systematic chromatographic analysis of antibiotics, of course, needs more detailed studies.

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SUMMARY

An attempt to establish a systematic chromatographic analysis of antibiotics has been made. Sixty-two known antibiotics were distributed into five classes and into fourteen subclasses, according to their R_F values in four principal solvent systems. Principles of the classification of antibiotics are given. For more detailed comparison and characterization of antibiotics further additional solvent systems were used. Additional solvent systems A, B, C, and D were used for class I, systems E, F, G, and H for classes II and III, systems I, J, K, and L were used for classes IV and V.

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